

# Novel Experimental Preparation to Assess Electrocardiographic Imaging Reconstruction Techniques

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## Abstract

*Electrocardiographic imaging (ECGI) systems are still plagued by a myriad of controllable and uncontrollable sources of error, which makes studying and improving these systems difficult. To mitigate these errors, we developed a novel experimental preparation using a rigid pericardiac cage suspended in a torso-shaped electrolytic tank. The 256-electrode cage was designed to record signals 0.5–1.0 cm above the entire epicardial surface of an isolated heart. The cage and heart were fixed in a 192-electrode torso tank filled with conductive fluid. The resulting signals served as ground truth for ECGI performed using the boundary element method (BEM) and method of fundamental solutions (MFS) with three regularization techniques: Tikhonov zero-order (Tik0), Tikhonov second-order (Tik2), truncated singular value decomposition (TSVD). Each ECGI regularization technique reconstructed cage potentials from recorded torso potentials well with spatial correlation above 0.7, temporal correlation above 0.8, and root mean squared error values below 0.7 mV. The earliest site of activation was best identified by MFS using Tik0, which localized it to within a range of 1.9 and 4.8 cm. Our novel experimental preparation has shown unprecedented agreement with simulations and represents a new standard for ECGI validation studies.*

## 1. Introduction

ECGI is an established technique that uses subject-specific geometry and body-surface electrocardiograms to reconstruct cardiac source signals. ECGI has high clinical applicability and has several commercial companies associated with its development. However, experimental and clinical validation studies for ECGI techniques have consistently shown significant errors from controllable and un-

controllable sources, including geometric registration, signal noise, conductivity, source modeling, and the inverse computation method.[1, 2] These various sources of error make it difficult to understand and interrogate ECGI approaches thoroughly, therefore there is a need for robust and controlled validation platforms with minimal sources of error.

To address this need, we developed a novel experimental preparation to limit sources of uncontrolled error. The preparation is similar to others used previously with a modified Langendorff perfused heart suspended inside a human shaped torso tank [3–5]. The torso tank allows us to control for several sources of error by utilizing a homogeneous isotropic volume conductor, recording high SNR body surface signals using electrodes in direct contact with the conductive media, and attaining expansive sampling using 192-electrodes spread across the tank surface. To further improve this preparation, we recently created a novel 256-electrode pericardiac cage electrode array to record signals immediately adjacent to the heart surface. The pericardiac cage array was rigidly fixed relative to the tank surface and sampled evenly around the entire cardiac source, further reducing geometric and source sampling errors. Using this model, we have shown significant improvements in the upper error bounds of the forward problem, and leveraged the sampling of the cardiac cage to investigate atrial signal reconstruction.[6, 7] However, we have not demonstrated the utility of the preparation for the inverse problem and ECGI.

In this report, we introduce our novel experimental preparation and demonstrate its value for validating the accuracy of ECGI-based reconstruction of ventricular-paced beats with a range of inverse techniques. Specifically, we examined an ECGI formulation defined by the boundary element formulation (BEM) of the forward problem and the method of fundamental solutions (MFS) with Tikhonov zero-order (Tik0), Tikhonov second-order (Tik2), and

truncated singular value decomposition (TSVD) regularization techniques. We reconstructed the cardiac potentials on the rigid pericardiac cage with ECGI and assessed each formulation using standard global metrics, signal-based comparisons, and clinically relevant standards.

## 2. Methods

**Experimental Preparation:** The experimental preparation was based on a modified Langendorff system using a support animal to sustain the normal function of an isolated heart.[3,5] In brief, a heart was isolated from a donor animal and connected via arterial and venous access to a support animal. The isolated heart was perfused through the coronary vasculature via the aorta and blood was returned to the support animal via a suction pump in the right ventricle. The isolated heart was attached to a rigid gantry, which could be precisely lowered and fixed in the torso tank. Registration points were recorded using a Microscribe digitizer throughout the experiment to mark the location of the recording electrodes. All experiments were performed under deep anesthesia. The procedures were approved by the Institutional Animal Care and Use Committee of the University of Utah and conformed to the Guide for the Care and Use of Laboratory Animals.

**Recording Arrays:** We used two electrical recording arrays in this study, as shown in Figure 1. The first was the novel cage array, which was constructed from 256 silver-silver chloride electrodes evenly spaced around the centroid of the cage. Spacing between each electrode location allowed electrical current to flow unimpeded to the torso tank surface. The cage was rigidly fixed around the entire heart 0.5–1.0 cm from the epicardial surface.

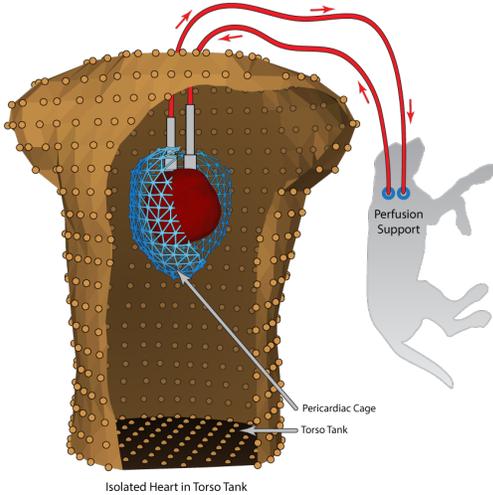


Figure 1. Schematic of experimental preparation with isolated heart, surrounded by the pericardiac cage and submerged in the torso tank.

The fixed cage and the Langendorff perfused heart were

then mounted in the torso tank, which has been used in previous studies and was modeled after a 10-year-old boy’s torso. The tank contains 192 embedded silver-silver chloride electrodes evenly spaced around the torso surface. The tank was filled with electrolytic fluid (500  $\Omega$  cm), which mimics the average conductivity of the human torso. Signals were acquired using a custom analog-to-digital multiplexer at 1 kHz with a bandpass filter at 0.3 to 500 Hz and a notch filter at 60 Hz. Signals were postprocessed, and annotated using methods described previously. [8]

**Beat Morphologies:** For this study, we selected beats recorded during anterior and posterior left ventricular free wall pacing using transmural plunge needle electrodes near the epicardial surface. For each site, 40 individual beats were identified, processed, and used to compare reconstruction techniques.

**Inverse Methods:** We applied four common inverse formulations to calculate cage potentials: MFS with Tik0 regularization and BEM with Tik0, Tik2, and TSVD regularization. These inverse methods were implemented in the SCIRun problem-solving environment using the Forward/Inverse Toolkit and MATLAB.[9] For the BEM regularizations, the L-curve corner was used to select the regularization parameter  $\lambda$  (Tik0 and Tik2) or the level of truncation (TSVD). For the MFS regularization with Tik0, we used the CRESO method to determine  $\lambda$ . The result was four inverse pipelines: BEM with Tik0, BEM with Tik2, BEM with TSVD, and MFS with Tik0.

**Evaluation Metrics:** To assess the accuracy of the inverse solutions, we calculated three standard statistical measures: RMSE, spatial correlation (SC), and temporal correlation (TC) according to the equations below. Here  $\Phi(k, n)$  is a  $K \times N$  matrix of measured ( $\Phi_m$ ) or calculated ( $\Phi_c$ ) potentials at  $K$  electrodes on the cage for  $N$  time instances. To calculate spatial or temporal correlation, we constructed zero mean vectors of either time signals on each electrode ( $\phi^t = \Phi(k, :) - \overline{\Phi(k, :)}$ ) or surface potential distributions at each time instance ( $\phi^s = \Phi(:, t) - \overline{\Phi(:, t)}$ ) respectively.

$$\text{RMSE} = \sqrt{\frac{\sum_{k=1}^K \sum_{n=1}^N ((\Phi_c(k, n) - \Phi_m(k, n))^2)}{N K}} \quad (1)$$

$$\text{SC} = \frac{1}{N} \sum_{n=1}^N \frac{\phi_m^s(n)^T \phi_c^s(n)}{jj\phi_m^s(n)jj jj\phi_c^s(n)jj} \quad (2)$$

$$\text{TC} = \frac{1}{K} \sum_{k=1}^K \frac{\phi_m^t(k)^T \phi_c^t(k)}{jj\phi_m^t(k)jj jj\phi_c^t(k)jj} \quad (3)$$

Previous studies have demonstrated that global statistics can be misleading.[1] To address this shortcoming, we implemented additional metrics: 1) the average magnitude of the spatial potential gradient at the peak of the RMS signal; 2) the activation localization error between the earliest

computed and measured activation sites, and 3) a spatial correlation of reconstructed and measured activation maps.

### 3. Results

Table 1 contains global statistics for all inverse approaches; the overall performances of most were comparable. Mean RMSE values, spatial correlation, and temporal correlation varied from 0.52–0.65 mV, 0.77–0.88, and 0.80–0.95, respectively. Across most global metrics, MFS with Tik0 performed consistently better with higher spatial and temporal correlations and slightly lower RMSE values. Correlations for activation maps ranged from 0.50–0.84, with MFS Tik0 performing best. Figure 2 contains samples of the best and worst inverse reconstructions with respect to spatial correlation.

The additional metrics also revealed differences in reconstruction performance. For example, the mean magnitudes of the measured gradient were 0.095–0.10 mV/mm and 0.073–0.065 mV/mm for the first and second ventricular-paced beat morphologies, respectively. BEM Tik0 and TSVD preserved the magnitude of the gradients better than BEM Tik2 or MFS Tik0. The localization error was slightly lower with BEM Tik0 and TSVD, both identifying the earliest site of activation equally with an mean range of 2.0–3.6 cm.

### 4. Discussion

In this study, we highlighted a new experimental preparation with a rigid pericardiac cage recording array and showed its utility as a validation platform for ECGI techniques. The preparation successfully attenuated many familiar sources of error, including geometric, signal noise, and torso conductivity. The recorded signals had both high resolution and coverage. We selected ventricular-paced beats for this initial assessment for their simplicity, wide use for validating ECGI, and their clinical relevance. We found that most of the inverse methods performed similarly based on global metrics, corroborating previous studies using simulated and experimental datasets. However, to our knowledge, the ECGI performance presented here was the highest published with an experimental validation model. These results must be carefully interpreted because the signals were recorded just above the heart surface and not directly on the epicardium. The cage possessed substantial advantages, such as uniform coverage over the entire heart, and its robustness to cardiac motion artifacts.

We assessed the reconstruction results with specialized, clinically driven metrics, for example an assessment of the average spatial gradient at the peak of the RMS. Accurate gradient reconstruction is essential to correctly identify the direction of wave propagation and early activation development. BEM Tik0 performed well at reconstructing the

gradient of potentials, which is expected because the Tik0 regularization smooths the signal based on amplitudes. In contrast, Tik2 smooths based on the spatial derivative and limits the gradients between neighboring sites. BEM Tik0 performed the best at localizing the earliest site of activation, even if this error was larger than that reported in other ECGI systems. We anticipated this result because of the spacing between the pericardiac cage and the epicardial surface. We did identify best case scenarios in which several of the experimental vs. computed locations were a perfect match. Despite the superior spatial and temporal correlation values of the MFS Tik0 technique, it did not perform well on the other signal or clinically driven metrics.

In summary, our experimental preparation has several unique and compelling features that we have designed to control various components of error related to the inverse problem and ECGI. We can examine the inverse problem in unprecedented detail in an environment in which many of the common errors plaguing ECGI are controlled. The rigid pericardiac cage geometry limits the amount of geometric error, and the homogeneous torso tank and embedded electrode arrays give us adequate sampling and high-quality signal recordings while simplifying the volume conductor models. We recognize that this preparation fails to completely represent human anatomy or physiology; however, the resulting datasets can be immensely valuable in testing fundamental characteristics related to ECGI and practical applications. Future studies will further refine the experimental model to perfect the validation platform and use it in a variety of ECGI and clinically driven contexts.

### Acknowledgments

Support for this research came from the NIH-NIGMS Center for Integrative Biomedical Computing ([www.sci.utah.edu/cibc](http://www.sci.utah.edu/cibc)), NIH NIGMS grant no.P41GM103545 and the Nora Eccles Treadwell Foundation for Cardiovascular Research.

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